

*The correspondence section is a public forum and, as such, is not peer-reviewed. EHP is not responsible for the accuracy, currency, or reliability of personal opinion expressed herein; it is the sole responsibility of the authors. EHP neither endorses nor disputes their published commentary.*

## Exposure to a Fungal Volatile Compound: Significance of Effects

In contrast to what Wälinder et al. (2005) concluded in their article "Acute Effects of a Fungal Volatile Compound," I interpret the article to report essentially no effects beyond chance. In all, the authors carried out some 76 comparisons (each one representing a time point and an exposure vs. control measurement) if you take blink frequency as a single comparison. The authors reported finding 5 "significant differences" out of 76 comparisons. Of the reported significant differences, one (blink frequency) is misleading, as discussed below. Of the remaining 75 comparisons, 4 differences at a  $p$ -value of  $< 0.05$  might be expected by random chance. This is without applying Bonferroni's adjustment for multiple comparisons; using this adjustment, a  $p$ -value of approximately  $< 0.0007$  would be required for a single comparison to be statistically significant. None of the differences reported reached this level.

Wälinder et al. (2005) reported that the subjects showed increased "blink frequency" during 3-methylfuran (3-MF) exposure (Table 1), but the frequency was higher in the exposure phase at time 0, about 9 for 3-MF exposure versus 6.5 for the control air phase. From the data in Table 1, it appears that both groups had fewer overall blinks per minute compared with baseline during the trial (Figure 2). Reporting that blinking was higher during exposure and not noting that it was higher at baseline is disingenuous.

Wälinder et al. (2005) may have mislabeled tear break-up, but as it reads in the legend for Table 1, a negative value indicates a decrease; therefore, the 6 sec given for measured break-up time after 3-MF exposure (Table 1) indicates that it was increased (i.e., longer to tear break-up), which is better. Is this correct? Also, was the observer who measured the tear break-up blinded to the exposure?

Finally, of the four lung measurements taken, the only comparison with a  $p$ -value of  $< 0.05$  was the small 100-mL change for forced vital capacity (FVC) right after exposure (Table 4). How do the authors interpret this change in FVC in view of the fact that there was no significant change in forced expiratory volume in 1 sec (FEV<sub>1</sub>)?

*The author has provided expert testimony in mold litigation.*

Andrew Saxon

Department of Medicine  
UCLA Medical School  
Los Angeles, California  
E-mail: asaxon@mednet.ucla.edu

## REFERENCE

Wälinder R, Ernstgård L, Johanson G, Norbäck D, Venge P, Wieslander G. 2005. Acute effects of a fungal volatile compound. *Environ Health Perspect* 113:1775–1778.

## Exposure to a Fungal Volatile Compound: Wälinder et al. Respond

There are different opinions on the use of Bonferroni's corrections. Everitt (1995) stated that it gives too conservative estimates if there are more than five tests performed. In environmental medicine, one exposure can have different health effects, so it is reasonable to test for different types of effects on different organs. We prefer to perform conventional statistical tests, without Bonferroni correction, and look at the pattern of significant effects and their biologic plausibility.

We did not perform 76 comparisons (Wälinder et al. 2005); we actually performed 25 tests on 13 physiologic variables based on differences before and after exposure. A fourteenth variable (vital staining) was tested only after exposure. Repeated measurement analysis was performed on blink frequency (60 consecutive measurements of 2 min each) and one questionnaire with 10 questions was administered at six different times. This is a total of 37 tests performed on 25 variables, having five significant values, of which one was highly significant ( $p < 0.001$ ).

Moreover, all tests point in the same direction—mucosal effects of the exposure. We did not find the same effects over time for control exposure. There is, of course, the possibility that some of the significant effects were due to chance, and we were quite modest in our conclusions, using the words "may" or "might be." Because our study is the first exposure-chamber study on 3-methylfuran (3-MF), more studies are needed to determine final conclusions.

Blink frequency was measured only during the 2 hr of exposure in the chamber. Therefore, there is no preexposure baseline data available at time 0. Eye effects occur quickly; a rapid effect of the exposure on blinking frequency can occur during the

first 2 min of exposure to 3-MF inside the chamber, possibly followed by later adaptation (8.8 blinks/min during the first 2 min, compared with a mean of 7.6 blinks/min during the whole period of exposure).

It is true that there was a numerical increase in break-up time at exposure, which could be in agreement with increased blink frequency. The fatty layer on the tear film is produced by the glands of the eyelids. Therefore, an increased blinking frequency could produce more secretion from the meibomian glands and therefore a longer break-up time.

Regarding lung function, transient effects of environmental exposure (as well as diurnal variation, which we controlled for by performing the experiment at the same time) may affect either forced vital capacity (FVC) or forced expiratory volume in 1 sec (FEV<sub>1</sub>), or both. Physiologically and numerically, the decrease was of the same order, but statistically the outcome was different. The decreases were 0.1 L for FVC and 0.08 L for FEV<sub>1</sub> after exposure to 3-MF. The magnitude of the effect was clinically small, but it was significant at group level for FVC. Small pulmonary effects may have large health effects in a population (Künzli et al. 2000.)

*The authors declare they have no competing financial interests.*

Robert Wälinder

Dan Norbäck

Gunilla Wieslander

Department of Medical Sciences/  
Occupational and Environmental Medicine  
University Hospital  
Uppsala, Sweden  
E-mail: robert.walinder@medsci.uu.se

Lena Ernstgård

Gunnar Johanson

Division of Work Environment Toxicology  
Institute of Environmental Medicine  
Karolinska Institutet  
Stockholm, Sweden

## REFERENCES

- Everitt BS. 1995. *The Cambridge Dictionary of Statistics in the Medical Sciences*. Cambridge, UK: Cambridge University Press.
- Künzli N, Ackermann-Liebrich U, Brändli O, Tschopp JM, Schindler C, Leuenberger P. 2000. Clinically "small" effects of air pollution on FVC have a large public health impact. *Eur Respir J* 15:131–136.
- Wälinder R, Ernstgård L, Johanson G, Norbäck D, Venge P, Wieslander G. 2005. Acute effects of a fungal volatile compound. *Environ Health Perspect* 113:1775–1778.

## Organophosphates and Outdoor Air Concentrations

Harnly et al. (2005) suggested that measured air concentrations of organophosphate insecticides may pose a particular concern for

children's health. However, when considering the current scientific weight of evidence, their conclusions cannot be supported for two reasons: first, they did not demonstrate a particular concern for children based on their results, and second, they cited incomplete and inappropriate literature to support the notion that recent toxicologic and epidemiologic studies indicate a health concern.

Harnly et al. (2005) did not conduct a risk assessment to demonstrate a concern for children. Rather, they detected pesticides in air concentrations in agricultural areas and then suggested there may be a concern for children because of recent toxicologic and epidemiologic studies. Their observed median exposures of all three active ingredients (chlorpyrifos, diazinon, and malathion) were all low and well within established regulatory limits. A risk assessment approach would have been quite useful. For chlorpyrifos, Harnly et al. (2005) detected a 20-day median concentration in air of 0.000033 mg/m<sup>3</sup>. A tier-1 risk assessment assuming an air concentration of chlorpyrifos at 0.000033 mg/m<sup>3</sup>, the mean body weight of a 1- to 2-year-old child of 12.3 kg, a child inhalation rate of 6.8 m<sup>3</sup>/day, and 24-hr outdoor respiration results in a chlorpyrifos inhalation exposure of 0.0000182 mg/kg/day. Margins of exposure (MOE) would be 5,495 [acute inhalation no observed adverse effect level (NOAEL) = 0.1 mg/kg/day], 27,473 (acute NOAEL = 0.5 mg/kg/day), and 1,648 (chronic NOAEL = 0.03 mg/kg/day). All MOEs are greater than the U.S. Environmental Protection Agency (EPA) target MOE of 1,000 for infants, children, and females 13–50 years of age (U.S. EPA 2002).

More problematic is that Harnly et al. (2005) stated that

Recent cellular, animal, and human evidence of toxicity, particularly in newborns, supports the public health concern indicated by initial risk estimates.

The authors did not provide a sufficiently thorough review of the literature relevant to risk assessment to support or refute their statement. In the case of chlorpyrifos, this statement cannot be supported by the available evidence. The principal problem with Harnly et al.'s approach is not unique to their article. Appropriate risk assessment requires appropriate data, and, as simple as this relationship sounds, it is often ignored. Harnly et al. (2005) cited toxicity and epidemiologic studies, but these particular studies are not appropriate for use in risk assessment. This problem has become so pervasive that Conolly et al. (1999) clarified the basic features of toxicology studies that are and are not appropriate for use in risk assessment.

Harnly et al. (2005) should not have included the findings of Qiao et al. (2002) because the high doses, subcutaneous route of administration, and carrier were inappropriate for toxicologic risk assessment (Conolly et al. 1999; Zhao et al. 2005). Indeed, Slotkin (2004), a coauthor of Qiao, has written that there is little academic interest in relevant routes of exposure or pharmacokinetics. He stated that

Practical issues that are critical to standardized testing are de-emphasized, such as pharmacokinetics/toxicokinetics, the matching of routes of exposure to those of humans in industrial, agricultural or domestic settings, or the development of biologically-based dose response models of established hazards. In that sense, the academic approach is entirely deficient in those attributes that are necessary components of the application of research findings to regulatory science.

Harnly et al. (2005) cited Eskenazi et al. (1999) as a source of concern for adverse consequences of organophosphate exposure. To be complete, Eskenazi et al. (2004) provide more information. They stated, "We failed to demonstrate an adverse relationship between fetal growth and any measure of *in utero* organophosphate pesticide exposure." An association was found for a couple of variables and decreased gestational duration, but the conclusion was that these potential pesticide effects appear to have "little clinical impact at the population level."

Finally, air concentrations have been shown to translate poorly into systemic exposure. Hore et al. (2005) showed that children in houses treated with chlorpyrifos had no detectable increase in urinary 3,5,6-trichloropyridinol (TCP), whereas median peak ambient air chlorpyrifos increased > 10-fold (median of 14 ng/m<sup>3</sup> pretreatment, 196 ng/m<sup>3</sup> on day of treatment). If a 10-fold increase in air chlorpyrifos does not cause a detectable increase in urinary TCP, then the 1-fold background air cannot be contributing measurably to the children's background levels of urinary TCP.

*The author was employed by Dow AgroSciences, a registrant of chlorpyrifos, from 1995 through 2001. Since he has been at Montana State University, he has received one unrestricted grant from Dow AgroSciences and one from The Dow Chemical Company to support his environmental risk assessment research. Additionally, he has had one research contract with Dow AgroSciences. None of the funding has involved research or other activities on chlorpyrifos.*

**Robert K.D. Peterson**

Agricultural and Biological Risk Assessment  
Department of Land Resources and  
Environmental Sciences  
Montana State University  
Bozeman, Montana  
E-mail: bpeterson@montana.edu

## REFERENCES

- Conolly RB, Beck BD, Goodman JI. 1999. Stimulating research to improve the scientific basis of risk assessment. *Toxicol Sci* 49:1–4.
- Eskenazi B, Bradman A, Castorina R. 1999. Exposures of children to organophosphate pesticides and their potential health effects. *Environ Health Perspect* 107(suppl 3):409–419.
- Eskenazi B, Harley K, Bradman A, Weltzien E, Jewell NP, Barr DB, et al. 2004. Association of *in utero* organophosphate pesticide exposure and fetal growth and length of gestation in an agricultural population. *Environ Health Perspect* 112:1116–1124.
- Harnly, M, McLaughlin R, Bradman A, Anderson M, Gunier R. 2005. Correlating agricultural use of organophosphates with outdoor air concentrations: a particular concern for children. *Environ Health Perspect* 113:1184–1189.
- Hore P, Robson M, Freeman N, Zhang J, Wartenberg D, Ozkaynak H, et al. 2005. Chlorpyrifos accumulation patterns for child-accessible surfaces and objects and urinary metabolite excretion by children for 2 weeks after crack-and-crevice application. *Environ Health Perspect* 113:211–219.
- Qiao D, Seidler FJ, Padilla S, Slotkin TA. 2002. Developmental neurotoxicity of chlorpyrifos: what is the vulnerable period? *Environ Health Perspect* 110:1097–1103.
- Slotkin TA. 2004. Guidelines for developmental neurotoxicity and their impact on organophosphate pesticides: a personal view from an academic perspective. *Neurotoxicology* 25(4):631–640.
- U.S. EPA. 2002. Interim Reregistration Eligibility Decision for Chlorpyrifos. EPA 738-R-01-007. Washington, DC: U.S. Environmental Protection Agency.
- Zhao Q, Gadagbui B, Douson M. 2005. Lower birth weight as a critical effect of chlorpyrifos: a comparison of human and animal data. *Regul Toxicol Pharm* 42:55–63.

## Organophosphates and Outdoor Air: Harnly et al. Respond

Peterson is incorrect in stating that we did not conduct a risk assessment. We summarized in our article (Harnly et al. 2005) and detailed in a previous article (Lee et al. 2002) a human health risk assessment demonstrating elevated acute and subchronic risks for children's exposures to ambient chlorpyrifos air concentrations in agricultural communities. Compared with the assessment by Peterson, we used a more refined probability distribution analysis, included air levels of the degradation product (chlorpyrifos oxon), and presented exposures relative to reference values. The chlorpyrifos reference values, however, were based on the same no-observed-adverse-effect levels (NOAELs) and the same 10-fold intraspecies and interspecies, and child uncertainty factors used in Peterson's calculations.

In our article (Harnly et al. 2005), we suggested that our risk assessment may underestimate risks for several reasons, such as *a*) risk assessments that use NOAELs, and not the entire dose–response curve, tend to underestimate risks (Castorina and Woodruff, 2003); and *b*) the true ranges of intraspecies and interspecies variability are unknown and may be larger than the factors used (Eskenazi et al. 1999; Faustmann et al. 2000). In a very recent study, some

newborns were 65–130 times less able to metabolize diazoxon and chlorpyrifos oxon than their mothers (Furlong et al. 2006). To further support the concern for children indicated by our quantitative risk assessment, we cited toxicologic studies establishing that in addition to cholinesterase inhibition, on which the NOAEL for chlorpyrifos is established, chlorpyrifos and chlorpyrifos oxon have other neurodevelopmental toxicity mechanisms (Huff et al. 1994; Qiao et al. 2002). We also noted that cell death has been induced at the reference dose for drinking water (Greenlee et al. 2005).

Peterson argues that the toxicologic studies we cited (Castorina and Woodruff, 2003; Eskenazi et al. 1999; Faustmann et al. 2000; Greenlee et al. 2005; Huff et al. 1994; Qiao et al. 2002) are an insufficient review of the “literature relevant to risk assessment” and that these studies are not appropriate for use in risk assessment. However, in missing the fact that we conducted a quantitative risk assessment, Peterson is misinterpreting our citations as the only basis for our public health concern. We consider it our public health responsibility to at least qualitatively consider recent toxicologic data in addition to a quantitative risk assessment based on established reference values. Others have argued for a complete restructuring of risk assessment for children, including toxicokinetic modeling and assessment of cellular and molecular outcomes over the entire lifespan of experimental subjects (Landrigan et al. 2004).

For many reasons we disagree with the suggestion that the epidemiologic fetal growth and gestational duration findings of Eskenazi et al. (2004) may be used to disregard concern for *in utero* and child organophosphate exposure highlighted by Eskenazi et al. (1999). The associations of reduced gestational duration with dimethyl organophosphate urinary metabolites and cholinesterase inhibition were not clinically significant in the California population studied (recent Mexican immigrants who tend to have very healthy birth outcomes). However, a shortened gestational age of a half-week would represent, for some women, a risk of preterm delivery (Eskenazi et al. 2004). Clearly, this finding and the absence of any adverse association between fetal growth and measures of *in utero* pesticide exposure need to be confirmed or refuted. To be complete, however, we also cited the association found in a New York City population between low birth weight and length and cord plasma levels of chlorpyrifos and diazinon ( $n = 314$ ) (Whyatt et al. 2004). Further, effects of organophosphate pesticide exposure on early child neurodevelopment have been found (Young

et al. 2005) and are continuing to be evaluated in the California and the New York City cohorts. Finally, public health policy is typically developed to protect against a 1 in 1,000, or lower, risk, and the epidemiologic studies cited here are below the sample size necessary to detect such risks.

Peterson notes that a study of children in 10 homes did not demonstrate an association with child urine metabolite levels of chlorpyrifos and ambient air levels following crack and crevice treatment (Hore et al. 2005). Yet, the authors of that study were careful to note a number of study limitations, including the variability and accuracy of the child urinary metabolite readings. We also note that chlorpyrifos oxon, which also breaks down into the measured urinary metabolite, was not measured in air; air concentrations in four of the study homes were not elevated compared to pretreatment levels; and personal air samples were not collected (Hore et al. 2005). Among mothers in New York City ( $n = 314$ ) in another study, 48-hr personal air samples collected during pregnancy were associated with cord and maternal blood levels of chlorpyrifos (Whyatt et al. 2004). This is the same study population within which an association with adverse birth outcomes and pesticide cord blood levels has been demonstrated, and the chlorpyrifos air levels are in the same (average, 15 ng/m<sup>3</sup>) range, if not lower, as those evaluated in our health risk assessment (Whyatt et al. 2004).

*The authors declare they have no competing financial interests*

**Martha Harnly  
Robert McLaughlin  
Robert Gunier**

California Department of Health Services  
Richmond, California  
E-mail: mharnly@dhs.ca.gov

**Asa Bradman**  
University of California  
Berkeley, CA

**Meredith Anderson**  
Impact Assessment Inc.  
Richmond, CA

## REFERENCES

- Castorina R, Woodruff TJ. 2003. Assessment of potential risk levels associated with U.S. Environmental Protection Agency reference values. *Environ Health Perspect* 111:1318–1325.
- Eskenazi B, Bradman A, Castorina R. 1999. Exposures of children to organophosphate pesticides and their potential health effects. *Environ Health Perspect* 107(suppl 3):409–419.
- Eskenazi B, Harley K, Bradman A, Weltzien E, Jewell NP, Barr DB, et al. 2004. Association of *in utero* organophosphate pesticide exposure and fetal growth and length of gestation in an agricultural population. *Environ Health Perspect* 112:1116–1124.
- Faustman EM, Silbernagel SM, Fenske RA, Burbacher TM,

- Ponce RA. 2000. Mechanisms underlying children's susceptibility to environmental toxicants. *Environ Health Perspect* 108(suppl 1):13–21.
- Furlong CE, Holland N, Richter RJ, Bradman A, Ho A, Eskenazi B. 2006. PON1 status of farmworker mothers and children as a predictor of organophosphate sensitivity. *Pharmacogenet Genomics* 16(3):183–190.
- Greenlee AR, Ellis TM, Berg RL. 2004. Low-dose agrochemicals and lawn-care pesticides induce developmental toxicity in murine preimplantation embryos. *Environ Health Perspect* 112:703–709.
- Harnly M, McLaughlin R, Bradman A, Anderson M, Gunier R. 2005. Correlating agricultural use of organophosphates with outdoor air concentrations: a particular concern for children. *Environ Health Perspect* 113:1184–1189.
- Hore P, Robson M, Freeman N, Zhang J, Wartenberg D, Ozkaynak H, et al. 2005. Chlorpyrifos accumulation patterns for child-accessible surfaces and objects and urinary metabolite excretion by children for 2 weeks after crack-and-crevice application. *Environ Health Perspect* 113:211–219.
- Huff RA, Corcoran JJ, Anderson JK, Abou-Donia MB. 1994. Chlorpyrifos oxon binds directly to muscarinic receptors and inhibits cAMP accumulation in rat striatum. *J Pharmacol Exp Ther* 269:329–335.
- Landrigan PJ, Kimmel CA, Correa A, Eskenazi B. 2004. Children's health and the environment: public health issues and challenges for risk assessment. *Environ Health Perspect* 112:257–265.
- Lee S, McLaughlin R, Harnly M, Gunier R, Kreutzer R. 2002. Community exposures to airborne agricultural pesticides in California: ranking of inhalation risks. *Environ Health Perspect* 110:1175–1184.
- Qiao D, Seidler FJ, Padilla S, Slotkin TA. 2002. Developmental neurotoxicity of chlorpyrifos: what is the vulnerable period? *Environ Health Perspect* 110:1097–1103.
- Whyatt RM, Rauh V, Barr DB, Camann DE, Andrews HF, Garfinkel R, et al. 2004. Prenatal insecticide exposures and birth weight and length among an urban minority cohort. *Environ Health Perspect* 112:1125–1132.
- Young JG, Eskenazi B, Gladstone EA, Bradman A, Pedersen L, Johnson C, et al. 2005. Association between *in utero* organophosphate pesticide exposure and abnormal reflexes in neonates. *Neurotoxicology* 26(2):199–209.

## Effects of BPA in Snails

It is an ethical requirement that new findings be presented in light of and in conjunction with a balanced evaluation of the current knowledge and published literature. We believe that Oehlmann et al. (2006) violated this general principle in several ways. For example, the authors inferred that prosobranch snails have a functional estrogen receptor and therefore a much higher sensitivity to estrogens and endocrine-disrupting compounds (EDCs) than other species previously reported in the literature. We found several other problems in their article:

First, Oehlmann et al. (2006) did not reveal the source of the animals used in their study, thus prohibiting independent repetition of the experiments by others.

Second, the authors stated that male and female *Marisa cornuarietis* cannot be distinguished morphologically without killing the animals. Therefore, the lack of data on the sex distribution of the animals sampled at each time-point leads us to question the stability of the experimental conditions with regard to sex ratios and thus reproductive conditions. Furthermore, the



rapidly changing snail density, and hence the sex distribution at each sampling time point, certainly influenced the remaining animals with respect to mortality and fecundity.

Third, the experimental design and the lack of replication (Experiment 1) did not allow for sound statistical analysis; the statistical methods used were inappropriate, making correct interpretation impossible. Of most concern to us was the analysis of data by analysis of covariance (ANCOVA), mainly because the ANCOVA-inherent assumption of independency of the dependent variable (i.e., total number of eggs) is violated. Thus, small differences among aquaria (treatment groups) might have been propagated over time, resulting in the impression of large differences.

Fourth, we believe that carrying out receptor binding experiments only in duplicate and without Scatchard analysis is questionable per se. The number of concentrations tested was extremely limited and consequently cannot allow accurate description of binding curves. Oehlmann et al. (2006) provided no information regarding the assessment of unspecific binding and the reported IC<sub>50</sub> values (concentration causing 50% inhibition) are approximately three orders of magnitude higher than what would be expected if this were a real sex-steroid receptor interaction. Because tamoxifen did not elicit a typical and highly specific receptor binding curve (Oehlmann et al. 2006, Figure 3), we question the use of tamoxifen as an "antiestrogen" in this *in vivo* study.

Finally, the data in Figure 1B (Oehlmann et al. 2006) were published earlier by Schulte-Oehlmann et al. (2001), yet the originally published data did not incorporate 17 $\alpha$ -ethinylestradiol (EE<sub>2</sub>) as positive control. Moreover, the EE<sub>2</sub> curve in Figure 1B appears identical to the one on slide 14 from a slide presentation available on Oehlmann's website (Schulte-Oehlmann et al. 2006).

The use of a positive control is commendable when the mode of action is known [National Toxicology Program (NTP) 2001]; however, as in the study of Oehlmann et al. (2006), the lack of such knowledge precludes the inclusion of a positive control as proof-of-principle. Slide 14 (Schulte-Oehlmann et al. 2006) demonstrates that EE<sub>2</sub> does not have a monotonic mode of activity in *M. cornuarietis*, but rather appears to stimulate egg laying at 10–25 ng EE<sub>2</sub>/L, inhibit egg laying at 50 ng EE<sub>2</sub>/L and has no effect at 1 and 100 ng EE<sub>2</sub>/L. On the basis of *in vitro* and *in vivo* effects reported by Oehlmann et al. (2006), we question the presence of any estrogen receptor-like interaction. In view of the NTP (2001) definitions and use of con-

trols, the use of EE<sub>2</sub> as a "positive" control, with its nonmonotonic and nonhormetic dose-response curve in comparison with BPA (which has a presumably monotonic response curve), as well as the use of an antiestrogen (tamoxifen), is inappropriate.

In conclusion, the data presented by Oehlmann et al. (2006) are unconvincing. Flaws in the experimental design, data presentation, and interpretation as well as statistical analyses render their findings untenable. Furthermore, the "Introduction" and "Discussion" of their article was written in a way that could be considered highly imbalanced and indeed alarmist. The highly selective inclusion/omission and discussion of previously published research that contradicts the authors' opinion (e.g., Pickford et al. 2003) is particularly disturbing. It is our opinion that our evaluation of the Oehlmann et al. work serves as a useful reminder to scientists that we must constantly strive to formulate clear hypotheses, use sound experimental designs, employ appropriate statistics, and draw conclusions that are supported by the available data and that reflect a balanced assessment of the scientific literature to avoid jumping to erroneous conclusions.

*The authors declare they have no competing financial interests.*

**Daniel R. Dietrich**  
**Evelyn O'Brien**

Human and Environmental Toxicology  
University of Konstanz  
Jacob-Burckhardstrasse, Germany  
E-mail: daniel.dietrich@uni-konstanz.de

**Sebastian Hoffmann**

European Commission  
Joint Research Centre  
Institute of Health and Consumer Protection  
Ispra, Italy

**Patrique Balaguer**  
**Jean-Claude Nicolas**

INSERM  
Endocrinologie Moléculaire et Cellulaire  
des Cancers  
Montpellier, France

**Willem Seinen**

Institute for Risk Assessment Sciences  
Utrecht, the Netherlands

**Michael Depledge**

Plymouth Marine Laboratories  
Plymouth, United Kingdom

## REFERENCES

- NTP. 2001. National Toxicology Program's Report of the Endocrine Disruptors Low Dose Peer Review. Research Triangle Park, NC:National Toxicology Program. Available: <http://ntp-server.niehs.nih.gov/ntp/htdocs/lowdosepeerfinalrpt.pdf> [accessed 14 April 2006].  
Oehlmann J, Schulte-Oehlmann U, Bachmann J, Oetken M, Lutz I, Kloas W, et al. 2006. Bisphenol A induces super-

feminization in the ramshorn snail *Marisa cornuarietis* (Gastropoda: Prosobranchia) at environmentally relevant concentrations. *Environ Health Perspect* 114(suppl 1): 127–133.

Pickford DB, Hetheridge MJ, Caunter JE, Hall AT, Hutchinson TH. 2003. Assessing chronic toxicity of bisphenol A to larvae of the African clawed frog (*Xenopus laevis*) in a flow-through exposure system. *Chemosphere* 53:223–235.

Schulte-Oehlmann U, Tillmann M, Casey D, Duft M, Markert B, Oehlmann J. 2001. Östrogenartige Wirkungen von Bisphenol A auf Vorderkiemensschnecken (Mollusca: Gastropoda: Prosobranchia). *UWSF Z Umweltchem Ökotoxikol* 13: 319–333.

Schulte-Oehlmann J, Nentwig G, Oetken M, Bachmann J, Oehlmann J. 2006. Effekte von ausgewählten Arzneimittelwirkstoffen auf aquatische Wirbellose. Available: [http://www.bio.uni-frankfurt.de/ee/ecotox/\\_files/teaching/hauptstudium/ecotox6.pdf](http://www.bio.uni-frankfurt.de/ee/ecotox/_files/teaching/hauptstudium/ecotox6.pdf) [accessed 18 April 2006].

## Effects of BPA in Snails: Oehlmann et al. Respond

We welcome critical appraisals that help to provide balance; however, Dietrich et al. gave an unjustified reproach. We feel that Dietrich's position is severely compromised because he serves as an expert for the bisphenol A (BPA) Industry Group (Brussels, Belgium). We would like to respond to the issues raised by Dietrich et al., as well as to their oversights and inappropriate interpretations of our findings.

The source of test animals was clearly provided in our "Materials and Methods" (Oehlmann et al. 2006). All animals were dissected and sexed; thus, sex distribution was known for each time-point of the experiment. We supposed a 1:1 sex ratio for dead snails, although historical data ( $n > 14,000$ ) indicate a slight prevalence of females (1.13:1); therefore, our assumption was conservative. Egg production was corrected for the number of females in the tanks, and snail densities were equal for all groups at each time-point.

Semistatic designs are widely applied in scientific and regulatory ecotoxicology [Organization for Economic Development and Co-operation (OECD) 1998]. The actual exposure concentrations of BPA were measured and clearly communicated in our Tables 1 and 2 (Oehlmann et al. 2006). Because 17 $\alpha$ -ethinylestradiol (EE<sub>2</sub>) is more stable than BPA (Larsson et al. 1999), exposure to the positive control is also guaranteed in our 24-hr renewal test. Interestingly, Dietrich himself coauthored a semistatic study on snails (Czech et al. 2001) with several shortcomings: they used no analytical verification of exposure concentrations, no replicates, and inconsistent group size.

Analysis of covariance (ANCOVA) analyses of fecundity, development, and other cumulative data are widely used (Bochdansky and Bollens 2004; Dziminski and Alford 2005; Schärer and Wedekind 1999). In our experiment 2 with replicates (Oehlmann et al. 2006), ANOVA confirmed the

ANCOVA results (Figure 2A,2C). A BPA Industry Group–sponsored statistical reevaluation of our raw data (Ecostat 2005) concluded that “at 20°C the mean egg production increased compared to the control in the BPA-exposed females at all applied concentrations (0.25, 0.5, 1 and 5 µg/L), and decreased in the BPA+faslodex- or tamoxifen-exposed females.”

We achieved an association for a steady state of specific binding in three independent time-course studies (Oehlmann et al. 2006). We determined nonspecific binding using a 1,000-fold excess of unlabeled ligands resulting in clear specific binding for testosterone and estradiol. At higher concentrations, nonspecific binding was 70%, comparable with findings of Chou and Dietrich (1999), who also performed their experiments in duplicate. This percentage might be due to homogenization of large amounts of tissue with high protein content but a limited degree of specific cytosolic binding sites. In our study (Oehlmann et al. 2006), we did not intend to deliver a complete binding study in which saturation experiments with Scatchard analysis are needed, but to provide indications for the presence of estrogen receptors by a specific binding of ligands to cytosolic extracts (a widely used practice). Tamoxifen was not disqualified as an antiestrogen because it elicited a binding higher than that of BPA. Furthermore, *in vitro* ligand affinities have a limited predictive value for biologic potencies *in vivo* (Kloas et al. 1999). In summary, the binding study was performed appropriately for the desired purpose and provides initial evidence for specific estrogen binding sites with high affinity for BPA.

Data presented in our Figure 1B (Oehlmann et al. 2006) were published in Schulte-Oehlmann et al. (2001) without EE<sub>2</sub> because the focus of that work was comparing responses to BPA in four prosobranch species, including *Marisa*. Because the article was published in German, the distribution was not large enough to bring the issue to a wider audience. In the current article (Oehlmann et al. 2006), EE<sub>2</sub> data were included to demonstrate the masking of BPA effects during the spawning season. Because future BPA industry-sponsored studies intend to investigate BPA effects under conditions maximizing reproduction, the problem of masked effects and an associated loss of sensitivity is of vital importance.

Responses in *Marisa* (ruptured oviducts, increased spawning) are estrogen specific and opposite of androgenic effects (imposex, reduced spawning). This and evidence communicated in our article (Oehlmann et al. 2006) justify the use of EE<sub>2</sub> to demonstrate the responsiveness of organisms. Non-

monotonic concentration responses have also been reported for estrogen-regulated end points in EE<sub>2</sub>-exposed fish (Pawlowski et al. 2004), supporting our view that estrogen-specific binding sites in *Marisa* may represent functional receptors.

Dietrich et al.'s charges that our “Introduction” and “Discussion” were “imbalanced and indeed alarmist” and that we selectively used literature are unjustified.

We hope that the evidence presented here serves to refute the unjustified claims made by Dietrich et al. We leave it to the readers to make final judgment, but we feel that with the ever-increasing body of evidence showing effects of BPA on reproduction in various animal species, common sense will eventually prevail on this issue.

*The authors declare they have no competing financial interests.*

**Jörg Oehlmann**

**Ulrike Schulte-Oehlmann**

**Matthias Oetken**

Johann Wolfgang Goethe University

Frankfurt am Main

Institute of Ecology, Evolution and Diversity

Frankfurt am Main, Germany

E-mail: oehlmann@zoology.uni-frankfurt.de

**Jean Bachmann**

Federal Environmental Agency

Section Ecological Assessment of

Substances

Dessau, Germany

**Ilka Lutz**

**Werner Kloas**

Leibniz-Institute of Freshwater Ecology

and Inland Fisheries

Department of Inland Fisheries

Berlin, Germany

**Thomas A. Ternes**

Federal Institute of Hydrology

Koblenz, Germany

## REFERENCES

- Bochdansky AB, Bollens SM. 2004. Relevant scales in zooplankton ecology: distribution, feeding, and reproduction of the copepod *Acartia hudsonica* in response to thin layers of the diatom *Skeletonema costatum*. *Limnol Oceanogr* 49:625–636.
- Chou YJ, Dietrich DR. 1999. Interactions of nitro-musk parent compounds and their amino-metabolites with the estrogen receptors of rainbow trout (*Oncorhynchus mykiss*) and the South African clawed frog (*Xenopus laevis*). *Toxicol Lett* 111: 27–36.
- Czech P, Weber K, Dietrich DR. 2001. Effects of endocrine modulating substances on reproduction in the hermaphroditic snail *Lymnaea stagnalis* L. *Aquat Toxicol* 53:103–114.
- Dziminski MA, Alford RA. 2005. Patterns and fitness consequences of intraclutch variation in egg provisioning in tropical Australian frogs. *Oecologia* 146:98–109.
- Ecostat. 2005. Experiments on the Effect of BPA on the Snail Species *Marisa cornuarietis* as Described in Three Papers by Oehlmann: Evaluation of the Applied Statistics and Analysis of the Raw Data. Report 05/011. Leiden:Ecostat—Statistical Consultancy in Ecology, Ecotoxicology and Agricultural Research.
- Kloas W, Lutz I, Einspanier R. 1999. Amphibians as a model to study endocrine disruptors: II. Estrogenic activity of environmental chemicals in vitro and in vivo. *Sci Total Environ* 225:59–68.
- Larsson DGJ, Adolfsson-Erici M, Parkkonen J, Pettersson M, Berg AH, Olsson PE, et al. 1999. Ethinylestradiol—an undesired fish contraceptive? *Aquatic Toxicol* 45:91–97.
- OECD. 1998. Guideline for Testing of Chemicals. *Daphnia magna* Reproduction Test. TG 211. Paris:Organization for Economic Co-operation and Development.
- Oehlmann J, Schulte-Oehlmann U, Bachmann J, Oetken M, Lutz I, Kloas W, et al. 2006. Bisphenol A induces superfeminization in the ramshorn snail *Marisa cornuarietis* (Gastropoda: Prosobranchia) at environmentally relevant concentrations. *Environ Health Perspect* 114(suppl 1): 127–133.
- Pawlowski S, van Aerle R, Tyler CR, Braunbeck T. 2004. Effects of 17 $\alpha$ -ethinylestradiol in a fathead minnow (*Pimephales promelas*) gonadal recrudescence assay. *Ecotoxicol Environ Saf* 57:330–345.
- Schärer L, Wedekind C. 1999. Lifetime reproductive output in a hermaphrodite cestode when reproducing alone or in pairs: a time cost of pairing. *Evol Ecol* 13:381–394.
- Schulte-Oehlmann U, Tillmann M, Casey D, Duft M, Markert B, Oehlmann J. 2001. Östrogenartige Wirkungen von Bisphenol A auf Vorderkiemerschnecken (Mollusca: Gastropoda: Prosobranchia). *UWSF Z Umweltchem Ökotoxikol* 13: 319–333.

## ERRATUM

In the article by Colbert et al. [*Environ Health Perspect* 113:700–707 (2005)], the authors discovered an error in the units of exposure estimates in the tenth paragraph of the “Discussion.” These estimates are expressed in milligrams but should be expressed in micrograms. The corrected sentences are as follows:

It is estimated that children 1–6 years of age are exposed to 0.167 µg Vz/kg body weight/day .... Given this chronic exposure estimate, a 2-year-old boy who weighs 13 kg ... would consume an average of 2.17 µg Vz/day, whereas a 6-year-old with a body weight of 21 kg would consume an average of 3.51 µg Vz/day.